SARS-CoV-2 Omicron And Multi-variant Neutralization Activity Of Ensovibep: A DARPin Therapeutic Candidate For Treatment Of Covid-19 Charles G. Knutson, Novartis Institutes for BioMedical Research, Cambridge, MA, USA

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Background

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- The omicron variant of SARS-CoV-2 has altered the COVID-19 pandemic landscape.
- Omicron's increased transmission and ability to evade natural or vaccine-induced immunity developed against earlier variants is a strong reminder of the power of viral evolution.
- Therapies with potential for multi-variant effectiveness are a key component of effective pandemic management.
- **ENSOVIBEP** is a first-in-class anti-SARS-CoV-2 DARPin (Designed Ankyrin Repeat Protein) therapeutic candidate that uses three distinct DARPin domains (R1, R2, R3) with similar paratopes to cooperatively bind to different regions of the receptor binding domain (RBD) of the SARS-CoV-2 spike protein trimer, thereby preventing interaction with the host ACE2 receptor.
- The multi-specific binding of the RBD binding DARPin modules limits the impact of spike protein mutations on antiviral potency (**Figure 1**).

Figure 1. Global frequencies of point mutations in the spike protein of SARS-CoV-2 according to the GISAID database, including a heat map table with IC₅₀ values for ensovibep, R1, R2, R3, for all point mutations tested.



Data from VSV/Lentivirus-based pseudovirus assays; dashed box: mutations in receptor binding domain.

- Depiction based on structural data showing ensovibep RBD binding DARPin domains (green, blue, cyan) binding to the RBD of the SARS-CoV-2 spike protein trimer. The two additional DARPin domains (purple) bind to human serum albumin (HSA, not shown for clarity) to provide half-life extension (Figure 2).
- We present here data supporting the multi-variant potency of ensovibep.
- Figure 2. Ensovibep bound to the SARS-CoV-2 spike protein.



HSA, human serum albumin; RBD, receptor binding domain.

Methods

- The VSV pseudotype viral system (CHUV) was based on the recombinant VSV*DELG-Luc vector in which the glycoprotein gene (G) was deleted and replaced with genes encoding green fluorescent protein and luciferase. Wild-type spike was based on the Wuhan-Hu-1 sequence. Pseudoviruses were mixed with serial dilutions of ensovibep and pre-incubated for 90 min at 37°C before adding to pre-seeded VeroE6 cells. After 90 min incubation, the inoculum was removed, fresh medium was added, and cells were further incubated for 16 h. Cells were lysed according to the ONE-Glo[™] luciferase assay system. Relative light units were measured and IC₅₀ values were calculated using nonlinear regression. Additional details can be found in Rothenberger et al 2022.
- Lentivirus pseudoviruses (ACTIV/FDA) bearing the spike proteins and carrying a firefly luciferase reporter gene were produced in 293T cells by co-transfection of pCMVDELR8.2, pHR'CMVLuc and pCDNA3.1(+)-spike variants. Wild-type spike was based on the Wuhan-Hu-1 sequence. Pseudoviruses were pre-incubated with serially diluted ensovibep for 2 h at 37°C before adding to pre-seeded 293T-ACE2.TMPRSS2s cells. Pseudovirus infection was scored 48 h later by measuring luciferase activity, and IC₅₀s were calculated using non-linear regression. Additional details can be found in Neerukonda et al 2021.
- Live authentic virus assay (Spiez Laboratory): Wild-type virus was a French isolate with the following changes compared with Wuhan-Hu-1: V367F, E990A. Serial dilutions of ensovibep were pre-incubated with 100 TCID₅0 SARS-CoV-2 variants for 1 h at 37°C before adding to pre-seeded VeroE6-TMPRSS2 cells. After 3 day incubation, cell viability was measured using CellTiter-Glo. Luminescence was measured and IC₅₀ values were calculated using non-linear regression. Additional details can be found in Rothenberger et al 2022.
- Live spike chimeric reporter viruses (UTMB) were constructed on the genetic background of an infectious cDNA clone derived from clinical strain WA1 (2019-nCoV/USA_ WA1/2020) containing a mNeoGreen (mNG) reporter gene, and spike mutations were engineered using a PCR-based mutagenesis protocol. The full-length genomic cDNAs were in vitro ligated, transcribed, and electroporated into VeroE6 cells, and mutant viruses were recovered 3 days after electroporation. Mutant viruses were pre-incubated with serial dilutions of ensovibvep for 1 h at 37°C before adding to pre-seeded Vero E6-TMPRSS2 cells. After 1 h infection, the inoculum was removed and replaced with overlay medium (DMEM with 0.8% methylcellulose, 2% FBS, and 1% P/S). After 16 h, raw images of mNG fluorescent foci were acquired, foci were counted, and IC₅₀s were determined using non-linear regression. Additional details can be found in Zou et al 2022.

Results

• Notably, ensovibep potency (IC₅₀) against all tested variants remains in the range of 1-10 ng/mL; less than an order of magnitude difference from the reference/Wuhan WT virus (Figure 3) (Rothenberger et al 2022).

Figure 3. Ensovibep activity measured in neutralization assays performed with Ientivirus, VSV-based pseudoviruses or authentic viruses for the SARS-CoV-2 variants of concern and variants of interest.



IC₅₀ (ng/ml)

Reference variant is the Wuhan-Hu-1 strain for VSV-based pseudovirus, a D614G variant for the lentivirus-based pseudovirus or a patient isolate from the early pandemic for the authentic virus. VSV, vesicular stomatitis virus.

Wild Type	Omicron BA.1		
IC ₅₀ (ng/mL)	IC ₅₀ (ng/mL)	Fold change to WT	
1.6	2.2	1.4	
3.2	>1000	>100	
3.3	>1000	>100	
13	>1000	>100	
6.4	>1000	>100	
23	72	3.1	
0.6	415	>100	
5.5	237	43	
9.5	392	41	
52	30	0.6	
	IC (ng/mL) 1.6 3.2 3.3 3.3 13 6.4 23 0.6 5.5 9.5 52 52	Wild TypeOmicroICICIC1.62.2 3.2 >1000 3.3 >100013>10006.4>100023720.64155.52379.53925230	

IC₅₀: green: <10 ng/mL; orange: 10-100 ng/mL; dark orange: 100-1000 ng/mL; red: >1000 ng/mL Fold change to wt: green: <10-fold; orange: 10–100-fold; red: >100-fold

Titration curves (right panels; mean \pm SEM) and IC₅₀ values (above) for VSV-pseudotype neutralization assays with wild-type and Omicron BA.1 variant spike protein. Ensovibep was tested together with a panel of clinically validated monoclonal antibodies. The table provides the numeric IC_{50} values as well as the fold change with respect to the wild-type. Wild-type virus is Wuhan-Hu-1



The table provides the numeric IC_{50} values as well as the fold change with respect to the wild-type. Wild-type virus is Wuhan-Hu-1.

(Figure 6).

Figure 6. Ensovibep activity against SARS-CoV-2 wild-type (Wuhan-Hu-1) and omicron BA.2 variant in VSVpseudotype neutralization assay.



The table provides the numeric IC_{50} values as well as the fold change with respect to the wild-type. Wild-type virus is Wuhan-Hu-1.

• Ensovibep was tested together with a panel of clinically relevant monoclonal antibodies (**Figure 4**). Figure 4. Neutralization activities (titration curves and IC₅₀) of ensovibep and monoclonal antibodies.



• A 10- to 40-fold increase in IC₅₀ was seen for each indvidual DARPin (R1, R2, R3), however the multispecific ensovibep containing all three RBD binding domains retained potency against BA.1 (Figure 5).

Figure 5. Titration curves and IC₅₀ values of individual ensovibep DARPin modules against wild-type and BA.1 variant of SARS-CoV-2 in VSV-pseudotype neutralization assay.

• Ensovibep maintained its potency against omicron BA.2, as seen from the titration curves and IC₅₀ values

Variant		IC ₅₀ (ng/mL) ^a		Fold change to
	Spike mutations	Wild type	Variant	Wild type
Delta / B.1.617.2	T19R, G142D, E156G, del157-158, L452R, T478K, D614G, P681R, D950N	3.8	3.0	0.8
Omicron / B.1.1.529 / BA.1	A67V, del69-70del, T95I, G142D, del143-145, del211, L212I, ins214EPE, G339D, S371L, S373P, S375F, K417N, N440K, G446S, S477N, T478K, E484A, Q493R, G496S, Q498R, N501Y, Y505H, T547K, D614G, H655Y, N679K, P681H, N764K, D796Y, N856K, Q954H, N969K, L981F	8.2	6.4	0.8
Omicron / B.1.1.529 / BA.2	T19I, del24-26, A27S, G142D, V213G, G339D, S371F, S373P, S375F, T376A, D405N, R408S, K417N, N440K, S477N, T478K, E484A, Q493R, Q498R, N501Y, Y505H, D614G, H655Y, N679K, P681H, N764K, D796Y, Q954H, N969K	3.8	12.1	3.2
Omicron / B.1.1.529 / BA.3	A67V, del69-70, T95I, G142D, del143-145, del211, L212I, G339D, S371F, S373P, S375F, D405N, K417N, N440K, G446S, S477N, T478K, E484A, Q493R, Q498R, N501Y, Y505H, D614G, H655Y, N679K, P681H, N764K, D796Y, Q954H, N969K	3.8	18.0	4.7
Omicron / B.1.1.529 / BA.2.12.1	T19I, del24-26, A27S, G142D, V213G, G339D, S371F, S373P, S375F, T376A, D405N, R408S, K417N, N440K, L452Q, S477N, T478K, E484A, Q493R, Q498R, N501Y, Y505H, D614G, H655Y, N679K, P681H, S704L, N764K, D796Y, Q954H, N969K	3.8 ^b	9.6	2.5
Omicron / B.1.1.529 / BA.4/5°	T19I, del24-26, A27S, del69-70, G142D, V213G, G339D, S371F, S373P, S375F, T376A, D405N, R408S, K417N, N440K, L452R, S477N, T478K, E484A, F486V, Q498R, N501Y, Y505H, D614G, H655Y, N679K, P681H, N764K, D796Y, Q954H, N969K	3.8 ^b	>10000	>2632

^aIC₅₀ values represent geometric means from 2–4 experiments. ^bWild-type experiments were not conducted in parallel.^cBA.4 and BA.5 share the same spike mutations. The table provides the numeric IC₅₀ values as well as the fold change with respect to the Wild-type virus is Wuhan-Hu-1

DMSO, Dimethyl sulfoxide

References

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Figure 7. Ensovibep activity against wild-type and omicron variants using live spike chimeric reporter virus.



• BA.4 and BA.5 share the same spike sequence, so one experiment was performed to represent both viruses. • Titration curves and IC₅₀ values for ensovibep against SARS-CoV-2 wild-type (WA-1) and omicron BA.2, BA.3, BA.2.12.1, BA.4/5, and delta variants showing potency is maintained against all variants assessed with the exception of BA.4/5 (**Figure 7**).

• BA.4/5 sub-lineages of the omicron variant have a F486V mutation that reduces binding of the RBD binding DARPin domains, which is consistent with prior results from RBD mutational analysis in pseudovirus systems (Figure 1).

Conclusions

• The neutralization potency of ensovibep is maintained across SARS-CoV-2 variants, including BA.1, BA.2 and BA.2.12.1, and BA.3 of the omicron sub-lineages.

 A reduction in neutralization potency was observed with omicron sub-lineages BA.4/5, which is likely attributed to the F486V mutation present in this variant. The global incidences of BA.4 and BA.5 is low (<5%), with the exception of South Africa and Portugal. The potential for BA.4 and BA.5 to increase in incidence is currently unknown.

• These findings highlight the multi-specific and cooperative binding characteristics of ensovibep, which was designed with the intent to develop a durable treatment that could continue to bind to the spike protein of a rapidly evolving virus.

Ensovibep continues to be investigated in clinical trials.

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