

EUROPEAN HEMATOLOGY ASSOCIATION

INTRODUCTION

- HSCT offers a potential cure for patients with AML and other malignant and non-malignant diseases. However:
- Many patients still relapse post-HSCT
- Current MAC is associated with toxicity which limits patient eligibility while RIC further increases the risk of relapse
- More effective and safe conditioning regimens are needed to improve outcomes in AML and allow broader access to HSCT in non-malignant diseases
- cKit (CD117), a critical receptor for stem cell maintenance, is an attractive target to selectively eliminate HSCs as well as LSCs in AML
- Preclinical studies in mice suggest that cKit-targeting Abs alone do not sufficiently deplete HSCs for efficient HSCT and would benefit from concomitant blockade of the CD47 "do-not-eat-me" signal¹
- However, α -CD47 Abs are limited by toxicities linked to CD47 expression on a wide range of healthy cells
- Hence, simply combining cKit-targeting and systemic CD47-blocking Abs for conditioning is not a viable approach in patients

AIM

• To achieve a safe and effective targeted conditioning by combining inhibition of c-Kit signaling with an active Fc-effector function and conditional blockade of CD47 on target cells in a single molecule to induce potent but selective depletion of c-Kit+ cells

METHOD

- Generation of MP0621, a cKit x CD16a x CD47 Switch-DARPin
- Containing a masked CD47-blocker that is released conditionally upon binding to cKit
- Triggering immune cell-mediated killing of HSCs by engaging macrophages and natural killer cells via CD16a



- Functional activity was assessed using macrophage phagocytosis assays
- The ability to conditionally block CD47 on cKit⁺ target cells was tested using CD47 competition and CD47/SIRPα inhibition in *in vitro* assays
- Serum half-life was assessed using mouse PK studies
- Efficacy to deplete cKit⁺ cells in BM was assessed using a hCD34⁺ humanized NSG mouse model

C-KIT x CD16a x CD47 Switch-DARPin With Conditional Blockade of CD47: A Next-Generation Targeted Conditioning For Hematopoietic Stem Cell Transplantation

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CONCLUSIONS

- Our preclinical results indicate that the c-KIT x CD16a x CD47 Switch-DARPin MP0621 can specifically target c-KIT⁺ HSCs with a suitable PK profile for a conditioning agent.
- The conditional CD47-binding enhances efficacy of cKit-targeting by blocking CD47 on target cells as well as safety by avoiding CD47-related off-target effects.
- Thus, MP0621 presents a novel targeted conditioning approach that has potential to improve the benefit/risk profile of HSCT strategies for patients with AML as well as non-malignant diseases.

PHARMACOKINETIC PROFILE OF MP0621 IS COMPATIBLE WITH HSCT CONDITIONING APPLICATION

- PK studies in mice indicate approx. mono-exponential decline of serum concentrations following single i.v. injection
- Half-life values in the mono-exponential phase were calculated to range from 2.5-3 h in mice which are expected to scale to approx. 0.5 d in human based on scaling with the half-life of HSA²







Figure 4. Treatment with MP0621 targets human cKit⁺ HSCs engrafted in CD34⁺ humanized NSG mice, while preventing elimination of peripheral hCD45⁺ cells observed with unconditional α -CD47 blockade. (A) Study design (B) Flow cytometry analysis of hCD45⁺ cells in peripheral blood before and after treatment with MP0621, α -cKit mAb (IgG1), or a combination of α -cKit mAb (IgG1) and α -CD47 (IgG4) mAbs. Displayed data are calculated as in the following equation: %hCD45+=(100*%hCD45+)/(%mCD45++%hCD45+); (Mann-Whitney test ****p<0.0001, ***p=0.007). (C-F) Flow cytometry assessment of human c-Kit+ cell populations in the bone marrow of hCD34-humanized NSG mice treated with MP0621 or α -cKit mAb vs vehicle (C) Representative flow cytometry dot plots and (D) frequency of human hCD34+ hcKit+ with HSC signature (hCD45^{dim} CD45RA⁻ HLADR⁺) (Kruskal-Wallis test ***p=0.004). (E) Representative flow cytometry plots and (F) frequency of BM hCD34⁻ hcKit⁺ cell population of total hCD45⁺ cells. (Kruskal-Wallis test **p=0.002; ***p=0.0004). Round and square symbols depict mice humanized with hCD34+HSCs isolated from cord blood of two different donors.

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REFERENCES

1. Chhabra et al., Sci Transl Med 2016;8(351); 2. Steiner et al., Protein Eng Des Sel. 2017 Sep 1;30(9):583-591; 3. Kwon et al., *Blood* 2019 May 9;133(19):2104-2108.

ABBREVIATIONS

mAb, (monoclonal) antibody; AML, acute myeloid leukemia; AUC, area under the curve; BM, bone marrow; ELISA, enzyme-linked immuno-sorbent assay E:T, effector to target; FACS, fluorescence-activated cell sorting; MAC, myeloablative conditioning; HSC(T), Hematopoietic stem cell (transplantation); HSA, human serum albumin; i.v., intravenous; LLOQ, lower limit of quantification; LSC, leukemic stem cell; MΦ, macrophage; PK, pharmacokinetic; RIC, reduced intensity conditioning; SIRPα, Signal regulatory protein α.

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Figure 3. PK characteristics MP0621 in female wild type BALB/c mice following single i.v. bolus injection of 1 and 9 mg/kg of MP0621 Results represent two groups with n=3 per time point (mean +/-max/min) Concentrations were measured by sandwich ELISA.

MP0621 TARGETS cKIT+ CELLS IN BONE MARROW WITHOUT

CONTACT INFORMATION

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